



Vivaldi Project

Data management plan

Crassostrea gigas response to OsHV-1 and vibrios (Harveyi clade)

Dual RNAseq

Oyster response to vibrios in the presence/absence of OsHV-1

04/02/2020

DATA MANAGEMENT PLAN

Template sheet for each dataset

| | |
|---|---|
| Partner name | CNRS |
| Data category | Transcriptome (host) |
| Concerned WP | WP2 |
| Name of the VIVALDI referent(s) | Transcriptome: Beatriz Novoa & Paola Venier |
| Reference of the dataset Please refer to the DMP table to find the appropriate reference. <i>Ex: Genome-Patho/SubTaskN*/Pathogen/PartnerN*</i> | Transcriptome-host/2.1.1/oyster/CNRS_IHPE |
| Description of the data | The response of oysters <i>C. gigas</i> to virulent/non-virulent vibrios was evaluated by RNAseq on oysters infected with a consortium of 18 strains of the Harveyi clade in the presence/absence of OsHV-1 virus. Material was one full-sib family of oysters susceptible to wild infections. The oyster and bacterial responses to infection were compared before mortalities occurrence and 4h, 24h and 48h after infection. |
| Type | Sequences |
| Period and frequency of data collection | Control and treated oysters sampled at 4, 24 and 48 h after infection |
| Geographical site of data collection (if applicable) | France |
| Description of the material from which the dataset is generated <i>Information will be obtained from individuals, which can come from natural/hatchery population and/or from family produced in hatchery. Animals can be infected (naturally or experimentally). DNA extraction can be done from the whole animal, tissue.</i> | <i>A biparental family of oysters (La Tremblade #H12). Whole tissues.</i> |
| Protocols <i>Example: 16S ribosomal RNA gene sequencing by NGS</i> Please refer to the DMP table* for more examples | <i>NGS sequencing of ribo-depleted RNA (Nugen technology) extracted from infected and non-infected oysters. Sequencing Novaseq 2x50 bp. FASTERis.</i> |
| Nature of the collected/generated data | fastq |



| | |
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| <p><i>Example: Raw dataset in .blc/.fastqc/.fasta formats for genomic information, and processed data set will be .vcf/.bed formats.</i> Please refer to the DMP table* for more examples</p> | |
| <p>Coverage (if applicable) <i>Example: random genomic regions covered at 50 X</i> Please refer to the DMP table* for more examples</p> | <p>100 millions paired-end reads per sample</p> |
| <p>What are the prerequisites allowing to use the data as such? <i>Example: Any person able to use .fastqc file and .fasta file</i> Please refer to the DMP table* for more examples</p> | <p>Any person able to use .fastqc file and .fasta file</p> |
| <p>Sharing of main data</p> | <p>Saved and shared after publication</p> |
| <p>Archiving and preservation <i>Example: data will be stored on a hard drive + online back up and then will be released on public database (Sinoe, Dryad) after publication.</i> Please refer to the DMP table* for more examples</p> | <p>SRA database</p> |
| <p>List, description and storage of associated data (metadata) <i>Examples: environmental data, mortality monitoring, genotyping...</i></p> | |
| <p>Sharing of metadata (if relevant)</p> | <p>No relevant</p> |