



Vivaldi Project

Data management plan

Responses of vibrios (Harveyi clade) to C. gigas colonization

Dual RNAseq

Bacterial response to oyster colonization in the presence/absence of OsHV-1 virus

30/05/2019

DATA MANAGEMENT PLAN

Template sheet for each dataset

Partner name	CNRS
Data category	Transcriptome (pathogen)
Concerned WP	WP2
Name of the VIVALDI referent(s)	Transcriptome: Beatriz Novoa & Paola Venier
Reference of the dataset Please refer to the DMP table to find the appropriate reference. <i>Ex: Genome-Patho/SubTaskN*/Pathogen/PartnerN*</i>	Transcriptome-pathogen/2.1.1/vibrio/CNRS_IHPE
Description of the data	<i>The response of vibrios to colonization was evaluated by RNAseq on oysters infected with 18 strains of the Harveyi clade (both virulent and non virulent) and in the presence/absence of OsHV-1. Material was one full-sib family of oysters susceptible to wild infections. The oyster and bacterial responses to infection were compared before mortalities occurrence at 0, 4, 24 and 48h after infection.</i>
Type	Sequences
Period and frequency of data collection	0, 4, 24 and 48h after infection.
Geographical site of data collection (if applicable)	n/a
Description of the material from which the dataset is generated <i>Information will be obtained from individuals, which can come from natural/hatchery population and/or from family produced in hatchery. Animals can be infected (naturally or experimentally). DNA extraction can be done from the whole animal, tissue.</i>	<i>A biparental family of oysters (La Tremblade H12)</i> Strain from the Harveyi clade <i>V. harveyi</i> O-A01, O-G11, O-D03 <i>V. rotiferianus</i> O-G05, O-E12, O-G10, O-B06, O-F05 <i>V. owensii</i> O-D04, Z-B04, F1-D04, O-A03, ZG08 <i>V. jasicida</i> F5-G08, F5-H11, F5-A12, F1-C08 <i>The strain RNAs were sequenced within the host tissues (whole tissue extracts)</i>
Protocols <i>Example: 16S ribosomal RNA gene sequencing by NGS</i> Please refer to the DMP table* for more examples	<i>NGS sequencing of ribo-depleted RNA (Nugen technology) extracted from infected and non-infected oysters. Sequencing Novaseq 2x50 bp. Fasteris.</i>
Nature of the collected/generated data	fastq



<p><i>Example: Raw dataset in .blc/.fastqc/.fasta formats for genomic information, and processed data set will be .vcf/.bed formats.</i></p> <p>Please refer to the DMP table* for more examples</p>	
<p>Coverage (if applicable)</p> <p><i>Example: random genomic regions covered at 50 X</i></p> <p>Please refer to the DMP table* for more examples</p>	<p>100 millions paired-end reads per sample</p>
<p>What are the prerequisites allowing to use the data as such?</p> <p><i>Example: Any person able to use .fastqc file and .fasta file</i></p> <p>Please refer to the DMP table* for more examples</p>	<p>Any person able to use .fastqc file and .fasta file</p>
<p>Sharing of main data</p>	<p>Saved and shared after publication</p>
<p>Archiving and preservation</p> <p><i>Example: data will be stored on a hard drive + online back up and then will be released on public database (Sinoe, Dryad) after publication.</i></p> <p>Please refer to the DMP table* for more examples</p>	<p>SRA database</p>
<p>List, description and storage of associated data (metadata)</p> <p><i>Examples: environmental data, mortality monitoring, genotyping...</i></p>	<p>Cliquez ici pour taper du texte.</p>
<p>Sharing of metadata (if relevant)</p>	<p>No relevant</p>

*To access the [DMP table](#), please login on the VIVALDI online platform

Once completed, this sheet has to:

1. Be sent to the referent(s) identified above for a final check
2. Be uploaded on the [VIVALDI online platform](#)

