



Vivaldi Project

Data management plan

BS-Seq, RNA-Seq – DNA methylation and RNA-Seq analysis on whole body oyster samples

30/03/2020

DATA MANAGEMENT PLAN

Template sheet for each dataset

Partner name	CNRS
Data category	Phenotypic markers (host)
Concerned WP	WP2 WP2 WP2
Name of the VIVALDI referent(s)	Phenotypic markers (host): Florian Enez & Christine Paillard
Reference of the dataset Please refer to the DMP table to find the appropriate reference. <i>Ex: Genome-Patho/SubTaskN*/Pathogen/PartnerN*</i>	Phenotypic-markers-host/2.1.4/C.Gigas/P2
Description of the data	The occurrence of epigenetic changes in <i>C. gigas</i> resisting to mortalities was studied in 2 full-sib families obtained from the DECIPHER project. The DNA methylation pattern (DNA methylation), gene expression (RNAseq) and phenotypic changes (increased resistance) in 2 generations (F1 and F2) of oysters submitted (or not) to immune stimulus (e.g. rich microbial environment) during ontogenesis or before the challenge was assessed.
Type	Sequences
Period and frequency of data collection	4 times: march 2017, July 2017, March 2018, July 2018
Geographical site of data collection (if applicable)	n/a
Description of the material from which the dataset is generated Information will be obtained from individuals, which can come from natural/hatchery population and/or from family produced in hatchery. Animals can be infected (naturally or experimentally). DNA extraction can be done from the whole animal, tissue.	A biparental oyster family (Decipher #11 and #32) DNA was extracted from whole body oysters sampled during larval development and juveniles.
Protocols Example: 16S ribosomal RNA gene sequencing by NGS Please refer to the DMP table* for more examples	DNA methylation data was generated from bisulfite converted gDNA sequenced by HiSeqX PE150bp (illumina) RNA data was generated by HiSeq4000 PE100bp (illumina)fastq



<p>Nature of the collected/generated data <i>Example: Raw dataset in .blc/.fastqc/.fasta formats for genomic information, and processed data set will be .vcf/.bed formats.</i> Please refer to the DMP table* for more examples</p>	<p><i>fastq</i></p>
<p>Coverage (if applicable) <i>Example: random genomic regions covered at 50 X</i> Please refer to the DMP table* for more examples</p>	<p><i>BS-Seq: 100-120 millions reads per sample RNA-Seq: 30 millions reads per samples</i></p>
<p>What are the prerequisites allowing to use the data as such? <i>Example: Any person able to use .fastqc file and .fasta file</i> Please refer to the DMP table* for more examples</p>	<p><i>Any person able to use .fastqc file</i></p>
<p>Sharing of main data</p>	<p><i>Saved and shared after publication Please specify</i></p>
<p>Archiving and preservation <i>Example: data will be stored on a hard drive + online back up and then will be released on public database (Sinoe, Dryad) after publication.</i> Please refer to the DMP table* for more examples</p>	<p><i>SRA database.</i></p>