Mucosal immunity in bivalves

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Rapid growth in immunity research parallels production growth



Significant progress

- Cellular and humoral processes of innate immunity
- Diversity of immune recognition factors (e.g. hypervariable immune proteins)
- Immune priming
- Role of the microbiome

Pending questions:

• What are the biological bases for resistance to infections





Immunity is provided by a multi-layered system

Only one particular site in animals encompasses all of these barriers:

MUCOSAL INTERFACES



Mucus: a great physical and biological barrier



Abundant mucosal secretions are produced by molluscan epithelial cells and subepithelial glands



- Can exceed 15% of energy gained from food (over 50% in some gastropods)
- Barrier to diffusion
- Selective ion transport

From Allam and Pales Espinosa, 2015 (Mucosal health in aquaculture, AP book)

Main mucosal epithelia in bivalve mollusks

Water processing organs

Major role of mucus in the transport of particles captured by the gills

Limited information on the role of bivalve mucus in immunity



pe: pallial epithelium ee: extrapallial epithelium ct: connective tissue wt: water tubule ij: interlamellar junction mf: muscle fibers sg: subepithelial gland ol: outer lobe pl: periostracal lamina dt: digestive tubule sd: secondary duct il: intestinal lumen

From Allam and Pales Espinosa, 2015



Bivalve pallial surfaces: hotspots for host-microbe interactions





Oysters filter >10 L h⁻¹ g⁻¹ dry tissue = over 25,000,000 microbes/second (considering a 1 g oyster and 10⁷ microbes ml⁻¹ seawater)

Pathogen acquisition mechanisms in marine bivalves

 Digestive tract (via mechanical and chemical activities) is an excellent barrier to infectious agents and most fatal infections affecting bivalve mollusks are initiated in pallial organs (mantle and gills)



QPX (clam mantle) *Vibrio tapetis* (clam mantle)

MSX (oyster gill)

Perkinsus marinus (oyster mantle)



Contents lists available at SciVerse ScienceDirect

(%)

Growth promotion or inhibition

Oyster mortality (%

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journal homepage: www.elsevier.com/locate/jip

Early host-pathogen interactions in marine bivalves: Evidence that the alveolate parasite *Perkinsus marinus* infects through the oyster mantle during rejection of pseudofeces

Bassem Allam ^{a,*}, Wade E. Carden ^{a,1}, J. Evan Ward ^b, Gina Ralph ^b, Sarah Winnicki ^{a,2}, Emmanuelle Pales Espinosa ^a

Vol. 104: 237-247, 2013 doi: 10.3354/dao02599	DISEASES OF AQUATIC ORGANISMS		
	Dis Aquat Org	Published June 13	

Early host-pathogen interactions in a marine bivalve: Crassostrea virginica pallial mucus modulates Perkinsus marinus growth and virulence

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Contents lists available at ScienceDirect

International Journal for Parasitology

journal homepage: www.elsevier.com/locate/ijpara

Pallial mucus of the oyster *Crassostrea virginica* regulates the expression of putative virulence genes of its pathogen *Perkinsus marinus* *

Emmanuelle Pales Espinosa^a, Erwan Corre^b, Bassem Allam^{a,*}

Significant regulation of virulence-related genes (RNASeq) in *P. marinus* following exposure to mucus





Results: RNASeq



Pales Espinosa et al. (2014). Int. J. Parasitology

RNASeq: some transcripts involved in interactions with host

Function	Up-regulated	Down-regulated		
Pro- apoptotic	14 transcripts : Apoptosis inducing factors LPS-induced TNF-α associated factor Macrophage migration inhibitory	6 transcripts: Apoptosis-inducing factor Programmed cell death		
Anti- apoptotic	11 transcripts: 14-3-3 proteins Apoptosis antagonizing transcription factor Fas apoptotic inhibitory	1 transcript: polyubiquitin		

• Manipulation of host cell apoptosis is an important trait of *P. marinus* infection in oyster (work by Sunila et al., Sokolova et al.)

• Parasite cyclophylins neutralize host antimicrobial peptides

Pales Espinosa et al. (2014). Int. J. Parasitology

What immune factors are present in mucus? Proteomic analysis (LC-MS/MS) of oyster mucus



Proteomic characterization of mucosal secretions in the eastern oyster, *Crassostrea virginica*

Emmanuelle Pales Espinosa ^{a,*}, Antonius Koller ^b, Bassem Allam ^a





What immune factors are present in mucus?

• Some proteins are differentially expressed in mucus derived from different organs



Mucus very rich in antimicrobial proteins including those involved in microbial binding (19 proteins: peptidoglycan-recognition protein, lectins) and killing (80 proteins: lysozyme, defensins)

Protein	Signal P	GenBank	eValue
C-type lectin-1 (CvML)	Signal P	BAF75353	1.97E-78
Galectin 2 (CvGal)	No	ABG75998	0
Macrophage mannose receptor 1	Extracellular/ Signal P	EKC26386	7.54E-64
Complement c1q-like proteins	Signal P	EKC25476	0
Thioester-containing proteins (TEPs)	Extracellular	EKC33672	0
Defensin	No	P85008	1.67E-18
Lysozyme 1	Extracellular/ Signal P	P83673	3.24E-116
Serine protease inhibitor (CvSI-2)	Extracellular/ Signal P	B9A8D7	1.01E-41

Molecular characterization of several mucosal lectins in bivalves

CvML (*Crassostrea virginica* mucocyte lectin) transcript levels (qPCR) 72 hrs after exposure to different treatments



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Some of these mucosal lectins interact with carbohydrates associated with microbial (microalgae) cell surface triggering their capture

Pales Espinosa et al., Biol Bull 2009, 2010a, 2010b; Jing et al., FSI 2011; Pales Espinosa and Allam, Mar Biol 2013

Mar Biol (2016) 163:40 DOI 10.1007/s00227-016-2815-0

ORIGINAL PAPER

Modeling food choice in the two suspension-feeding bivalves, Crassostrea virginica and Mytilus edulis

Emmanuelle Pales Espinosa¹ · Robert M. Cerrato¹ · Gary H. Wikfors² · Bassem Allam¹

Oysters and mussels preferentially ingest microalgae having mannose residues on their cell surface







RESEARCH ARTICLE

Reverse genetics demonstrate the role of mucosal C-type lectins in food particle selection in the oyster *Crassostrea virginica*

Emmanuelle Pales Espinosa* and Bassem Allam



Mucus covering pallial surfaces also contains blood cells (hemocytes)





Fish and Shellfish Immunology

journal homepage: www.elsevier.com/locate/fsi

Full length article

Characterization of hemocytes from different body fluids of the eastern oyster *Crassostrea virginica*

Yuk-Ting Lau, Lauren Sussman, Emmanuelle Pales Espinosa, Selma Katalay¹, Bassem Allam*



Journal of Invertebrate Pathology

journal homepage: www.elsevier.com/locate/jip

Transepithelial migration of mucosal hemocytes in *Crassostrea virginica* and potential role in *Perkinsus marinus* pathogenesis

Yuk-Ting Lau, Laura Gambino, Bianca Santos, Emmanuelle Pales Espinosa, Bassem Allam*



Fish and Shellfish Immunology

journal homepage: www.elsevier.com/locate/fsi

Full length article

Regulation of oyster (*Crassostrea virginica*) hemocyte motility by the intracellular parasite *Perkinsus marinus*: A possible mechanism for host infection

Yuk-Ting Lau, Laura Gambino, Bianca Santos, Emmanuelle Pales Espinosa, Bassem Allam®



Fish and Shellfish Immunology

journal homepage: www.elsevier.com/locate/fsi

Full length article

Regulation of apoptosis-related genes during interactions between oyster hemocytes and the alveolate parasite *Perkinsus marinus*

Yuk-Ting Lau, Bianca Santos, Michelle Barbosa, Emmanuelle Pales Espinosa, Bassem Allam*

Conclusions

- Mucosal interfaces contain a wide range of immune factors
- Specific host-microbe interactions at these interfaces are determinant for some of the most basic physiological processes in bivalves
- Some of these non-self recognition factors have been diverted for the optimization of energy gains
- Hemocytes associated with mucosal surfaces occupy a niche similar to that occupied by dendritic cells in vertebrates

Acknowledgements



- Co-PI and co-author: Emmanuelle Pales Espinosa
- Funding: National Science Foundation
- Many students, in particular Joyce Lau, Wade Carden, Sarah Winnicki, Michelle Barbosa and Anna Hollembeak





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Exchange exists between circulatory and pallial hemocytes



Tracking of hemolymph hemocytes. Fluorescent hemocytes peaked in both compartments at day 1. N=11 to 16 oysters/data point.



Lau et al., in preparation

Pallial hemocytes can also reach the circulatory system



Tracking of pallial hemocytes. Fluorescent hemocytes were visible in the circulatory system on day 1 and peaked after 48 hours. N=10 oysters/ data point.



Lau et al., in preparation

Oyster exposure to *Perkinsus marinus* increases hemocyte trafficking



New question: can the parasite take advantage of hemocyte trafficking to get access into host tissues?









P. marinus 42um under mantle surface

В



P. marinus cells can be internalized by pallial hemocytes and transferred to hemolymph











