



# Vivaldi Project

## Data management plan

**Genome (pathogen)**

**Key words:** OsHV-1, *Crassostrea gigas*, Illumina sequencing

07/08/2018

## DATA MANAGEMENT PLAN

### Template sheet for each dataset

<b>Partner name</b>	UNIPD
<b>Data category</b>	Genome (pathogen)
<b>Concerned WP</b>	WP1 WP2 WP3
<b>Name of the VIVALDI referent(s)</b>	Genome (OsHV-1): Benjamin
<b>Reference of the dataset</b> <i>Please refer to the DMP table to find the appropriate reference.</i> <small>Ex: Genome-Patho/SubTaskN*/Pathogen/PartnerN*</small>	Genome-Patho/subtask 1.2.1/Pathogen/P12
<b>Description of the data</b>	Whole genome sequence of the viral isolate OsHV-1-PT (PT means virus isolated from <i>C. gigas</i> farmed in the Porto Tolle area)
<b>Type</b>	Sequences
<b>Period and frequency of data collection</b>	Oysters sampled during late Spring 2016
<b>Geographical site of data collection (if applicable)</b>	North Adriatic Sea, Po river delta, P. Tolle area
<b>Description of the material from which the dataset is generated</b> <i>Information will be obtained from individuals, which can come from natural/hatchery population and/or from family produced in hatchery. Animals can be infected (naturally or experimentally). DNA extraction can be done from the whole animal, tissue.</i>	The infection propagation trial started from a tissue homogenate supernatant obtained from farmed oysters resulted to be OsHV-1 positive: -total DNA from gills/mantle tissue fragments of oysters ( $\leq 4$ -5 months age, Porto Tolle area, North Adriatic Sea, Italy); -q-RT-PCR diagnosis with the HVDP-F /HVDP-R primers targeting the catalytic subunit of the viral DNA polymerase: ORF100, nucleotides 147655-153291 of the reference OsHV-1 genome (AY509253); - $2.5$ - $2.8 \times 10^8$ virus DNA copies/100 $\mu$ l (see Tab. 1 Abbadi et al. 2018).
<b>Protocols</b> <i>Example: 16S ribosomal RNA gene sequencing by NGS</i> <b>Please refer to the DMP table* for more examples</b>	All protocols, including whole genome shotgun sequencing and de novo assembly, are detailed in Abbadi et al. 2018 (DOI 10.1099/jgv.0.001042). See the technical summary at NCBI (PRJNA398466 and SRS2433575).
<b>Nature of the collected/generated</b>	About the OsHV-1-PT genome, raw data are in the SRA archive under

<p><b>data</b> <i>Example: Raw dataset in .blc/.fastqc/.fasta formats for genomic information, and processed data set will be .vcf/.bed formats.</i> <b>Please refer to the DMP table* for more examples</b></p>	<p>accession number SRR5936879 whereas the consensus sequence is recorded in GenBank under accession number MG561751). The OsHV-1-PT genome organization, equal to that of OsHV-1 <math>\mu</math>Var, can be represented as TR<sub>L</sub>-U<sub>L</sub>-IR<sub>L</sub>-X-IR<sub>S</sub>-U<sub>S</sub>-TR<sub>S</sub>-X' or X'-TR<sub>L</sub>-U<sub>L</sub>-IR<sub>L</sub>-X-IR<sub>S</sub>-U<sub>S</sub>-TR<sub>S</sub>.</p>
<p><b>Coverage (if applicable)</b> <i>Example: random genomic regions covered at 50 X</i> <b>Please refer to the DMP table* for more examples</b></p>	<p><b>200-fold base pair sequence coverage</b> and a <b>279-fold physical coverage</b> of the OsHV-1 genome. Illumina sequencing was directly performed on a virus-rich oyster sample and yielded 3 436 820 paired-end reads (2x300 bp), which allowed the recovery of 87 582 high-quality reads truly belonging to the order Herpesvirales (2.6 % OsHV-1 DNA to exogenous DNA ratio).</p>
<p><b>What are the prerequisites allowing to use the data as such?</b> <i>Example: Any person able to use .fastqc file and .fasta file</i> <b>Please refer to the DMP table* for more examples</b></p>	<p>Anyone interested in the molecular OsHV-1-oyster interactions and in the mitigation of OsHV-1-related pathogenicity</p>
<p><b>Sharing of main data</b></p>	<p><i>Saved and shared after publication</i> <i>Please specify</i></p>
<p><b>Archiving and preservation</b> <i>Example: data will be stored on a hard drive + online back up and then will be released on public database (Sinoe, Dryad) after publication.</i> <b>Please refer to the DMP table* for more examples</b></p>	<p><i>Please refer to the public repositories SRA and GenBank as they represent a reference for the whole scientific community.</i></p>
<p><b>List, description and storage of associated data (metadata)</b> <i>Examples: environmental data, mortality monitoring, genotyping...</i></p>	<p>See SSampling table to Fill-in.xls (IZSVE16_PT OsHV1 pos CG) See publicly available data (SRR5936879 and MG561751) See Abbadi et al 2018 (DOI 10.1099/jgv.0.001042)</p>
<p><b>Sharing of metadata (if relevant)</b></p>	<p><i>Choisissez un élément.</i> <i>Please specify</i></p>

\*To access the [DMP table](#), please login on the VIVALDI online platform

Once completed, this sheet has to:

1. Be sent to the referent(s) identified above for a final check
2. Be uploaded on the [VIVALDI online platform](#)

